



## Rapid Standardized Laboratory Protocol for Molecular Subtyping of *Vibrio parahaemolyticus* by Pulsed Field Gel Electrophoresis (PFGE)

### PREPARATION OF PFGE PLUGS FROM AGAR CULTURES

**BIOSAFETY WARNING:** *Vibrio parahaemolyticus* is a human pathogen that can cause serious disease. Always use Biosafety Level 2 practices and extreme caution when transferring and handling strains of this genus. Work in a biological safety cabinet when handling large amounts of cells. Disinfect or dispose of all plasticware and glassware that come in contact with the cultures in a safe manner.

Please read all instructions carefully before starting protocol. Treat all plasticware, glassware, pipets, spatulas, etc. that come in contact with the cell suspensions or plugs as contaminated materials and dispose of, or disinfect according to the guidelines of your institution. Disinfect reusable plug molds before they are washed; the disposable plug molds, including the tape and the tab that is used to push the plugs out of the wells, are also contaminated and should be disinfected with 10% bleach for at least 30 minutes if they will be washed and reused.

#### Day 0

Streak an isolated colony from test cultures to Trypticase Soy Agar with 5% defibrinated sheep blood (TSA-SB) plates (or comparable media) for confluent growth. It is recommended that a storage vial of each culture be created. To do this stab small screw cap tubes of TSA, HIA, or similar medium with the same inoculating loop used to streak the plate. This will ensure that the same colony can be retested if necessary. Incubate cultures at 37°C for 14-18 h.

#### Day 1

1. **Turn on shaker water bath (54-55°C), stationary water baths (55-60°C) and spectrophotometer** (or equivalent instrument such as the Dade Microscan Turbidity meter or bioMérieux Vitek colorimeter).

2. Prepare **TE Buffer (10 mM Tris:1 mM EDTA, pH 8.0)**<sup>1</sup> as follows:

10 ml of 1 M Tris, pH 8.0

2 ml of 0.5 M EDTA, pH 8.0

Dilute to 1000 ml with sterile Ultrapure water (Clinical Laboratory Reagent Water (CLRW))

**Note:** The TE Buffer is used to make the plug agarose and also to wash lysed PFGE plugs.

3. Prepare 1% SeaKem Gold agarose in **TE Buffer (10 mM Tris:1 mM EDTA, pH 8.0)**<sup>2</sup> for PFGE plugs as follows:

a. Weigh 0.50 g (or 0.25 g) SeaKem Gold (SKG) into a 250 ml screw-cap flask.

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<sup>1</sup>Additional information is found on page 14 or in Section 5a of the PulseNet PFGE Manual.

- b. Add 49.5 ml (or 24.75 ml – 25 ml) TE Buffer; swirl gently to disperse agarose.
- c. Remove cap, cover loosely with clear film, and microwave for 30-sec; mix gently and repeat for 10-sec intervals until agarose is completely dissolved.
- d. Place melted agarose in a 55- 60°C water bath for 15 minutes or until ready to use.

**SAFETY WARNING:** Use heat-resistant gloves when handling hot flasks after microwaving.

**Note:** SeaKem Gold agarose works well for making PFGE plugs because it provides added strength to the plugs, minimizing breakage of plugs during the lysis and washing steps. The time and temperature needed to completely dissolve the agarose is dependent on the specifications of the microwave used and will have to be determined empirically in each laboratory.

4. Label small tubes (12-mm x 75-mm Falcon tubes or equivalent) with culture numbers.
5. Prepare **Cell Suspension Buffer (100 mM Tris:100 mM EDTA, pH 8.0)** as follows:

10 ml of 1 M Tris, pH 8.0  
20 ml of 0.5 M EDTA, pH 8.0  
Dilute to 100 ml with sterile Ultrapure water (CLRW)

6. Transfer ≈2 ml of **Cell Suspension Buffer (CSB)** to small labeled tubes. Use a sterile polyester-fiber or cotton swab that has been moistened with sterile CSB to remove some of the growth from agar plate; suspend cells in CSB by spinning swab gently so cells will be evenly dispersed and formation of aerosols is minimized.

**Note:** The minimum volume of the cell suspension needed will depend on size of the cuvettes or tubes used to measure the cell concentration and are dependent on the manufacturer's specifications for the spectrophotometer, turbidity meter, or colorimeter.

7. Adjust concentration of cell suspensions to one of values given below by diluting with sterile CSB or by adding additional cells.
  - a. Spectrophotometer: 610 nm wavelength, absorbance (Optical Density) of 0.9 (range of 0.8-1.0)
  - b. Dade Microscan Turbidity Meter: **0.35 - 0.45** (measured in Falcon 2054 tubes)  
**0.55 – 0.65**(measured in Falcon 2057 tubes)
  - c. bioMérieux Vitek colorimeter: ≈**20%** transmittance (measured in Falcon 2054 tubes)

**Note:** The values in Steps 7a, 7b and 7c give satisfactory results at CDC; each laboratory may need to establish the optimal concentration needed for satisfactory results.

## CASTING PLUGS

Label wells of PFGE plug molds with culture number. When reusable plug molds are used, put strip of tape on lower part of reusable plug mold before labeling wells.

**Note 1:** Unused plug agarose can be kept at room temperature and reused 1-2 times. Microwave on low-medium power for 10 -15 sec and mix; repeat for 5 -10 sec intervals until agarose is completely melted. This agarose melts rapidly!

**Note 2:** Proteinase K solutions (20 mg/ml) are available commercially, or a stock solution of Proteinase K can be prepared from the powder in sterile Ultrapure water (CLRW), aliquoted in 300-500  $\mu$ l amounts, and kept frozen. Just before use, thaw appropriate number of vials needed for the samples; keep Proteinase K solutions on ice. If the Proteinase K stock solution was prepared from powder, discard any thawed solution at the end of work day. Store commercially prepared Proteinase K solutions according to directions provided by the supplier.

1. Transfer 400  $\mu$ l (0.4 ml) adjusted cell suspensions to labeled 1.5-ml microcentrifuge tubes.
2. Add 20  $\mu$ l of Proteinase K (20 mg/ml stock) to each tube and mix gently with pipet tip. (200  $\mu$ l are needed for 10 cell suspensions.)
3. Add 400  $\mu$ l (0.4 ml) melted 1% SeaKem Gold agarose<sup>2</sup> to the 0.4-ml cell suspension; mix by gently pipetting mixture up and down a few times. Maintain temperature of melted agarose by keeping flask in beaker of warm water (55-60°C).
4. Immediately, dispense part of mixture into appropriate well(s) of reusable plug mold. Do not allow bubbles to form. Two plugs of each sample can be made from these amounts of cell suspension and agarose and are useful if repeat testing is required. Allow plugs to solidify at room temperature for 10-15 min. They can also be placed in the refrigerator (4°C) for 5 minutes.

**Note:** If disposable plug molds are used for making plugs with 1% SeaKem Gold agarose, use 200  $\mu$ l cell suspension, 10  $\mu$ l of Proteinase K (20 mg/ml stock) and 200  $\mu$ l of agarose; up to 4 plugs can be made from these amounts of cell suspension and agarose.

**Note:** The generation of cell suspension and the subsequent casting of the plugs should be performed as rapidly as possible in order to minimize premature cell lysis. If large numbers of samples are being prepared, it is recommended that they be processed in batches of ~10 samples at a time. Once the first batch of isolates are in the cell lysis incubation, then start preparing the cells suspensions the next group samples, and so on. All batches can be lysed and washed together, since additional lysis time will not affect the initial batches.

## LYSIS OF CELLS IN AGAROSE PLUGS

**Note:** Two plugs (reusable plug molds) or 3 - 4 plugs (disposable plug molds) of the same strain can be lysed in the same 50-ml tube.

1. Label 50-ml polypropylene screw-cap or 50-ml Oak Ridge tubes with culture numbers.
2. Prepare **Cell Lysis Buffer (50 mM Tris:50 mM EDTA, pH 8.0 + 1% Sarcosyl)** as follows:
  - 25 ml of 1 M Tris, pH 8.0
  - 50 ml of 0.5 M EDTA, pH 8.0
  - 50 ml of 10 % Sarcosyl (N-Lauroylsarcosine, Sodium salt)<sup>3</sup>
  - Dilute to 500 ml with sterile Ultrapure water (CLRW)
3. Calculate the total volume of **Cell Lysis/Proteinase K Buffer** needed as follows:
  - a. 5 ml **Cell Lysis Buffer (50 mM Tris:50 mM EDTA, pH 8.0 + 1% Sarcosyl)** is needed per tube (e. g., 5 ml x 10 tubes = 50 ml).
  - b. 25 µl **Proteinase K** stock solution (20 mg/ml) is needed per tube of the cell lysis buffer (e. g., 25 µl x 10 tubes = 250 µl).
  - c. Prepare the master mix by measuring the correct volume of Cell Lysis Buffer and Proteinase K into appropriate size test tube or flask and mix well.

**Note:** The final concentration of Proteinase K in the lysis buffer is 0.1 mg/ml, and is different from the concentration that was added to the cell suspension (0.5 mg/ml).

4. Add 5 ml of Proteinase K/Cell Lysis Buffer to each labeled 50 ml tube.
5. Trim excess agarose from top of plugs with scalpel or razor blade (optional). Open reusable plug mold and transfer plugs from mold with a 6-mm wide spatula to appropriately labeled tube. If disposable plug molds are used, remove white tape from bottom of mold and push out plug(s) into appropriately labeled tube. Be sure plugs are under buffer and not on side of tube.

**Note:** The excess agarose, plug mold, spatula, etc. are contaminated. Discard or disinfect appropriately.

6. **Remove tape from reusable mold.** Place both sections of plug mold, spatulas, and scalpel in 70% isopropanol (IPA) or other suitable disinfectant. **Soak them for 15 minutes before washing them.** Discard disposable plug molds or disinfect them in 10% bleach for 30-60 minutes if they will be washed and reused.
7. Place tubes in rack and incubate in a 54-55°C shaker water bath for **1 hour** with constant and vigorous agitation (150-175 rpm). Be sure the water level in water bath is above the level of the lysis buffer in the tubes.

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<sup>3</sup>The N-Lauroylsarcosine, Sodium salt can be added directly to the other ingredients and allowed to dissolve. See page 14 of this document or Section 5a of the PulseNet PFGE Manual.

8. Pre-heat enough sterile Ultrapure water (CLRW) to 54-55°C so that plugs can be washed two times with 10-15 ml water (200-250 ml for 10 tubes).

### WASHING OF AGAROSE PLUGS AFTER CELL LYSIS

**Note:** Most laboratories will find that their plugs are sufficiently stable to perform the following washing steps at 54-55°C. However, if you notice that your plugs are nicked along the edges or breaking it will be necessary for your laboratory to lower the water bath or incubator to 50°C for the following washing steps.

1. Remove tubes from water bath, and carefully pour off lysis buffer into an appropriate discard container; plugs can be held in tubes with a screened cap or spatula.

**Note:** It is important to remove all of the liquid during this and subsequent wash steps by touching edge of tube or screened cap on an absorbent paper towel.

2. Add at 10-15 ml sterile Ultrapure water (CLRW) that has been pre-heated to 54-55°C to each tube and shake the tubes vigorously in a 54-55°C water bath for 10-15 min.
3. Pour off water from the plugs and repeat wash step with pre-heated water (Step 2) one more time.
  - a. Pre-heat enough sterile **TE Buffer (10 mM Tris:1 mM EDTA, pH 8.0)** in a 54-55°C water bath so that plugs can be washed four times with 10-15 ml TE (300-350 ml for 10 tubes) after beginning last water wash.
4. Pour off water, add 10-15 ml pre-heated (54-55°C) sterile TE Buffer, and shake the tubes vigorously in 55°C water bath for 10-15 min.
5. Pour off TE and repeat wash step with pre-heated TE three more times.
6. Decant last wash and add 5-10 ml sterile TE. Continue with step 1 (Restriction Digestion" section or store plugs in TE Buffer at 4°C until needed. Plugs can be transferred to smaller tubes for long term storage.

**Note:** If restriction digestion is to be done the same day, complete Steps 1-3 of next section (Restriction Digestion) during last TE wash step for optimal use of time.

### RESTRICTION DIGESTION OF DNA IN AGAROSE PLUGS WITH *Sfi*I or *Not*I

**Note:** A small slice of the plug or the entire plug (made in disposable plug molds) can be digested with the restriction enzyme. Restriction digestion of a small slice of the plug is recommended because less enzyme is required and other slices of the plug can be subjected to restriction analysis with other enzymes. This is important when the PFGE patterns obtained with the primary enzyme from two or more isolates are indistinguishable, and confirmation is needed to determine that the PFGE patterns of these isolates are also indistinguishable with additional enzymes.

1. Label 1.5-ml microcentrifuge tubes with culture numbers; label 3 (10-well gel) or 4 (15-well gel) tubes for *Salmonella* ser. Braenderup H9812<sup>4</sup> standards.

**Note:** The appropriate restriction buffer will vary between vendors and may differ between enzymes from the same vendor. Always use the restriction buffer recommended by the vendor for the particular restriction enzyme.

- a. **Optional Pre-Restriction Incubation Step:** Prepare a master mix by diluting the appropriate 10X restriction buffer (Roche Molecular Biochemicals or equivalent) 1:10 with sterile Ultrapure water (CLRW) according to the following table.

Reagent	µl/Plug Slice	µl/7 Plug Slices	µl/11 Plug Slices
<b>Sterile Clinical Laboratory Reagent Water</b>	180 µl	1260 µl	1980 µl
<b>Restriction Buffer</b>	20 µl	140 µl	220 µl
<b>Total Volume</b>	200 µl	1400 µl	2200 µl

- b. Add 200 µl diluted restriction buffer (1X) to labeled 1.5-ml microcentrifuge tubes.
- c. Carefully remove plug from TE with spatula and place in a sterile disposable Petri dish or on large glass slide.
- d. Cut a 2.0- to 2.5-mm-wide slice from each test samples and the appropriate number of *S.* ser. Braenderup H9812 standard with a single edge razor blade (or scalpel, cover slip, etc.) and transfer to tube containing diluted restriction buffer. Be sure plug slice is under buffer. Replace rest of plug in original tube that contains 5 ml TE buffer and store at 4°C.

**Note:** The shape and size of the plug slice that is cut will depend on the size of the comb teeth that are used for casting the gel. PulseNet recommends that the combs with larger teeth (10-mm-wide teeth) be used to cast the gels because computer analysis of the gel lanes is more accurate and less tedious than analysis of gel lanes cast with combs with the smaller teeth (5.5-mm). The number of slices that can be cut from the plugs will depend on the skill and experience of the operator, integrity of the plug, and whether the slices are cut vertically or horizontally (plugs made in disposable molds).

- f. Incubate sample and control plug slices in 50°C (*SfiI*) or 37°C (*NotI* and *XbaI*) water bath for 5-10 min or at room temperature for 10-15 min.
- g. After incubation, remove buffer from plug slice using a pipet fitted with 200-250 µl tip all the way to bottom of tube and aspirate buffer. Be careful not to cut plug slice with pipet tip and that plug slice is not discarded with pipet tip.

<sup>4</sup> Directions for making and testing PFGE plugs of *Salmonella* ser. Braenderup H9812 are in the PulseNet QA/QC Manual

2. Prepare the restriction enzyme master mix by diluting 10X restriction buffer 1:10 with sterile Ultrapure water (CLRW), BSA, and *Sfi*I, *Not*I, or *Xba*I (*S. Braenderup* H9812 – see footnote 4 on page 5) restriction enzymes according to the following tables. Mix in the same tube that was used for the diluted the restriction buffer.

**Note:** The *Sfi*I stock enzyme should be ordered in concentrated form (40 U/ $\mu$ l) rather than unconcentrated form (10 U/ $\mu$ l). The unconcentrated form is acceptable for *Not*I restriction of *V. parahaemolyticus*.

Reagent	$\mu$ l/Plug Slice	$\mu$ l/7 Plug Slices	$\mu$ l/11 Plug Slices
<b>Sterile Clinical Laboratory Reagent Water</b>	176.75 $\mu$ l	1237.25 $\mu$ l	1944.25 $\mu$ l
<b>Restriction Buffer</b>	20 $\mu$ l	140 $\mu$ l	220 $\mu$ l
<b>BSA (10mg/ml)</b>	2 $\mu$ l	14 $\mu$ l	22 $\mu$ l
<b><i>Sfi</i>I (40 U/<math>\mu</math>l)</b>	1.25 $\mu$ l	8.75 $\mu$ l	13.75 $\mu$ l
<b>Total Volume</b>	200 $\mu$ l	1400 $\mu$ l	2200 $\mu$ l

Reagent	$\mu$ l/Plug Slice	$\mu$ l/7 Plug Slices	$\mu$ l/11 Plug Slices
<b>Sterile Clinical Laboratory Reagent Water</b>	174 $\mu$ l	1218 $\mu$ l	1914 $\mu$ l
<b>Restriction Buffer</b>	20 $\mu$ l	140 $\mu$ l	220 $\mu$ l
<b>BSA (10mg/ml)</b>	2 $\mu$ l	14 $\mu$ l	22 $\mu$ l
<b><i>Not</i>I (10 U/<math>\mu</math>l)</b>	4 $\mu$ l	28 $\mu$ l	44 $\mu$ l
<b>Total Volume</b>	200 $\mu$ l	1400 $\mu$ l	2200 $\mu$ l

**Note:** Keep vial of restriction enzyme on ice or in insulated storage box (-20°C) at all times.

3. Add 200  $\mu$ l restriction enzyme mixture to each tube. Close tube and mix by tapping gently; confirm that plug slices are under enzyme mixture.
4. Incubate sample and standard (control) plug slices for 4 hours in a water bath at the appropriate temperature for the enzyme.
- Incubate samples restricted with *Sfi*I at 50°C
  - Incubate samples restricted with *Not*I and *Xba*I at 37°C.
5. If plug slices will be loaded into the wells (Option B, page 9), continue with Steps 1-4 of the next section (**CASTING AGAROSE GEL**) approximately 1 h before restriction digest reaction is finished so the gel can solidify for at least 30 minutes before loading the restricted PFGE plugs.

## CASTING AGAROSE GEL

### A. Loading Restricted Plug Slices on the Comb:

1. **Confirm that water bath is equilibrated to 55- 60°C.**
2. Make volume of 0.5X Tris-Borate EDTA Buffer (TBE) that is needed for both the gel and electrophoresis running buffer according to one of the following tables.

#### 5X TBE:

Reagent	Volume in milliliters (ml)					
<b>5X TBE</b>	200	210	220	230	240	250
<b>Reagent Grade Water<sup>5</sup></b>	1800	1890	1980	2070	2160	2250
<b>Total Volume of 0.5X TBE</b>	2000	2100	2200	2300	2400	2500

#### 10X TBE:

Reagent	Volume in milliliters (ml)					
<b>10X TBE</b>	100	105	110	115	120	125
<b>Reagent Grade Water</b>	1900	1995	2090	2185	2280	2375
<b>Total Volume of 0.5X TBE</b>	2000	2100	2200	2300	2400	2500

3. Make 1% SeaKem Gold (SKG) Agarose in 0.5X TBE as follows:
  - a. Weigh appropriate amount of SKG into 500 ml screw-cap flask.
  - b. Add appropriate amount of 0.5X TBE; swirl gently to disperse agarose.
  - c. Remove cap, cover loosely with clear film, and microwave for 60-sec; mix gently and repeat for 15-sec intervals until agarose is completely dissolved.
  - d. Recap flask and place in 55-60°C water bath for 15 minutes or until ready to use.

Mix 1.0 g agarose with 100 ml 0.5X TBE for 14-cm-wide gel form (10 or 15 wells)

Mix 1.5 g agarose with 150 ml 0.5X TBE for 21-cm-wide gel form (≥15 wells)

**SAFETY WARNING:** Use heat-resistant gloves when handling hot flasks after microwaving.

4. A small volume (2-5 ml) of melted and cooled (55-60°C) 1% SKG agarose may be wanted to seal wells after plugs are loaded. Prepare 50 ml by melting 0.5 g agarose with 50 ml 0.5X TBE in 250 ml screw-cap flask as described above. Unused SKG agarose can be kept at room temperature, melted, and reused several times. Microwave for 15-20 sec and mix; repeat for 10-sec intervals until agarose is completely melted. Place in 55-60°C water bath until ready to use. Alternatively, 3-5 ml of the melted agarose used to cast the gel can be transferred to a pre-heated (55-60°C) 50 ml flask and kept in 55-60°C water bath and used to seal the wells..

<sup>5</sup> De-ionized water (does not need to be sterilized).



**Note:** Confirm that gel form is level on leveling table, that **front** of comb holder and teeth face the bottom of gel, and that the comb teeth touch the gel platform.

5. Remove restricted plug slices from 50°C or 37°C water bath. Remove enzyme/buffer mixture and add 200 µl 0.5X TBE. Incubate at room temperature for 5 min.
6. Remove plug slices from tubes; put comb on bench top and load plug slices on the bottom of the comb teeth as follows:
  - a. Load *S. ser.* Braenderup H9812 standards on teeth (lanes) 1, 5, 10 (10-well gel) or on teeth 1, 5, 10, 15 (15-well gel).
  - b. Load samples on remaining teeth.
7. Remove excess buffer with tissue. Allow plug slices to air dry on the comb for ≈5 minutes or seal them to the comb with 1% SKG agarose (55-60°C).
8. Position comb in gel form and confirm that the plugs slices are correctly aligned on the bottom of the comb teeth, that the lower edge of the plug slice is flush against the black platform, and there are no bubbles (if allowed to air dry).
9. Carefully pour the agarose (cooled to 55-60°C) into the gel form.
10. Put black gel frame in electrophoresis chamber. Add 2 -2.2 L freshly prepared 0.5X TBE. Close cover of unit. (The amount of buffer needed depends on whether residual buffer was left in tubing or if the unit was flushed with water after the last gel was run.)
11. Turn on cooling module (14°C), power supply, and pump (setting of ≈70 for a flow of 1 liter/minute).
12. Remove comb after gel solidifies for 30-45 minutes.
13. Fill in wells of gel with melted and cooled (55-60°C) 1% SKG Agarose (optional). Unscrew and remove end gates from gel form; remove excess agarose from sides and bottom of casting platform with a tissue. Keep gel on casting platform and carefully place gel inside black gel frame in electrophoresis chamber. Close cover of chamber.

#### **B. Loading Restricted Plug Slices into the Wells:**

1. Follow steps 1-4 in Option A on pages 7 and 8 (**Loading Restricted Plug Slices on the Comb**).

**Note:** Confirm that gel form is level on gel-leveling table before pouring gel, that front of comb holder and teeth face bottom of gel, and the bottom of the comb is 2 -mm above the surface of the gel platform.

2. Cool melted SKG agarose in 55-60°C water bath for 15-20 min; carefully pour agarose into gel form (casting stand) fitted with comb. Be sure there are no bubbles.

3. Put black gel frame in electrophoresis chamber. Add 2-2.2 L freshly prepared 0.5X TBE. Close cover of unit. (The amount of buffer depends on whether residual buffer was left in tubing, or if unit was flushed with water after the last gel was run.)
4. Turn on cooling module (14°C), power supply, and pump (setting of  $\approx 70$  for a flow of 1 liter/minute) approximately 30 min before gel is to be run.
5. Remove restricted plug slices from 50°C or 37°C water bath. Remove enzyme/buffer mixture and add 200  $\mu$ l 0.5X TBE. Incubate at room temperature for 5 minutes.
6. Remove comb after gel solidifies for at least 30 minutes.
7. Remove restricted plug slices from tubes with tapered end of spatula and load into appropriate wells. Gently push plugs to bottom and front of wells with wide end of spatula. Manipulate position with spatula and be sure that are no bubbles.
  - a. Load *S. ser.* Braenderup H9812 standards in wells (lanes) 1, 5, 10 (10-well gel) or in wells 1, 5, 10, 15 (15-well gel).
  - b. Load samples in remaining wells.

**Note:** Loading the plug slices can be tedious; each person has to develop his/her own technique for consistently placing the plug slices in the wells so the lanes will be straight and the bands sharp.

8. Fill in wells of gel with melted 1% SKG Agarose (equilibrated to 55- 60°C). Allow to harden for 3-5 min. Unscrew and remove end gates from gel form; remove excess agarose from sides and bottom of casting platform with a tissue. Keep gel on casting platform and carefully place gel inside black gel frame in electrophoresis chamber. Close cover of chamber.

### ELECTROPHORESIS CONDITIONS

1. Program the CHEF Mapper with the following switch conditions for *V. parahaemolyticus* strains restricted with *Sfi*I and *Not*I:
  - Auto Algorithm
  - 78 kb - low MW
  - 396 kb - high MW
  - Select default values except where noted by pressing "enter".
  - Change run time to 18hr** (See note below)
  - (Default values: Initial switch time = 10 s; Final switch time = 35.03 s)
  - linear ramping factor

Select the following conditions on **CHEF DR-III**

- Initial A time: 10 s
- Final A time: 35 s
- Start ratio: 1.0
- Voltage: 200 V
- Run time: 18 hr

**Note:** The electrophoresis running times recommended above are based on the equipment and reagents used at the CDC. Run times may be different in your laboratory and will have to be optimized for your gels so that the lowest band in the *S. ser. Braenderup H9812* standard migrates 1.0 - 1.5 cm from the bottom of the gel.

**Note:** Make note of the initial milliamp (mA) reading on the instrument. The initial mA should be between 110-170 mA. A reading outside of this range may indicate that the 0.5X TBE buffer was prepared improperly and the buffer should be remade.

## Day 2

### STAINING AND DOCUMENTATION OF PFGE AGAROSE GEL

1. When the electrophoresis run is over, turn off equipment; remove and stain gel with ethidium bromide. Dilute 40 µl of ethidium bromide stock solution (10 mg/ml) with 400 ml of reagent grade water (this volume is for a staining box that is approximately 14-cm x 24-cm; a larger container may require a larger amount of staining solution). Stain gel for 20 - 30 min in covered container.

**Note:** Ethidium bromide is toxic and a mutagen; the solution can be kept in dark bottle and reused 4 - 5 times before discarding according to your institution's guidelines for hazardous waste or use the destaining bags recommended for disposal of ethidium bromide (**Section 10**).

2. Destain gel in approximately 500 ml reagent grade water for 60 - 90 min; change water every 20 minutes. Capture image on Gel Doc 1000, Gel Doc 2000, or equivalent documentation system. If background interferes with resolution, destain for an additional 30-60 min.

**Note:** If both a digital image and conventional photograph are wanted, photograph gel first before capturing digital image.

3. Follow directions given with the imaging equipment to save gel image as an \*.img or \*.1sc file; convert this file to \*.tif file for analysis with the BioNumerics software program. The gel image should fill the entire window of the imaging equipment (computer) screen (without cutting off wells or lower bands). Ensure that the image is in focus and that there is little to no saturation (over-exposure) in the bands. Additional instructions are provided in PNL07 of the PulseNet QA/QC manual.
4. Drain buffer from electrophoresis chamber and discard. Rinse chamber with 2 L reagent grade water or, if unit is not going to be used for several days, flush lines with water by letting pump run for 5-10 min before draining water from chamber.
5. If the lowest band in the H9812 standard does not migrate within 1-1.5 cm of the bottom of the gel, the run time will need to be determined empirically for the conditions in each laboratory.

**Please note the following if PFGE results do not have to be available within 24-28 hours:**

1. Plugs can be lysed for longer periods of time (3-16 hours).

2. The washing steps with TE to remove the lysis buffer from the PFGE plugs can be done for longer periods of time (30-45 min) and at lower temperatures (37°C or room temperature). They can be started on Day 1 and finished on Day 2 after overnight refrigeration of the plugs in TE.
3. The restriction digestion can be done for longer periods of time (3-16 hours).
4. If the lowest band in the H9812 standard does not migrate within 1 -1.5 cm of the bottom of the gel, the run time will need to be determined empirically for the conditions in each laboratory.

Use of trade names and commercial sources is for identification purposes only and does not imply endorsement by CDC or the U.S. Department of Health and Human Services.

**NOTE: CLIA LABORATORY PROCEDURE MANUAL REQUIREMENTS**

Efforts have been made to assure that the procedures described in this protocol have been written in accordance with the 1988 Clinical Laboratory Improvement Amendments (CLIA) requirements for a procedure manual (42 CFR 493.1211). However, due to the format required for training, the procedures will require some modifications and additions to customize them for your particular laboratory operation.

Any questions regarding the CLIA requirements for a procedure manual, quality control, quality assurance, etc., should be directed to the agency or accreditation organization responsible for performing your laboratory's CLIA inspection. In addition, some states and accreditation organizations may have more stringent requirements that will need to be addressed.



**Formulas of Selected Reagents used in PulseNet Standardized Laboratory Protocol for PFGE**

**Tris:EDTA Buffer, pH 8.0 (TE, 10 mM Tris:1 mM EDTA, pH 8.0)<sup>6</sup>**

10 ml of 1 M Tris, pH 8.0  
2 ml of 0.5 M EDTA, pH 8.0  
Dilute to 1000 ml with sterile Ultrapure water (CLRW)

**Cell Lysis Buffer (50 mM Tris:50 mM EDTA, pH 8.0 + 1% Sarcosine + 0.1 mg/ml Proteinase K)**

25 ml (50 ml) of 1 M Tris, pH 8.0  
50 ml (100 ml) of 0.5 M EDTA, pH 8.0  
50 ml (100 ml) 10% N-Lauroylsarcosine, Sodium salt (Sarcosyl)

**or**

5 g (10 g) of N-Lauroylsarcosine, Sodium salt (Sarcosyl)<sup>7</sup>  
Dilute to 500 ml (1000 ml) with Sterile Ultrapure water (CLRW)

Add 25 µl Proteinase K stock solution (20 mg/ml) per 5 ml of cell lysis buffer just before use for a final concentration in the lysis buffer of 0.1 mg/ml Proteinase K.

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<sup>6</sup>This formula for TE is from Molecular Cloning - A Laboratory Manual by J. Sambrook and E. Russell, 3<sup>rd</sup> edition. TE Buffer from Life Technologies (CP0558; 0126A) used at CDC is 0.01M (10 mM) for both ingredients. To duplicate this commercial formula, increase the amount of 0.5 M EDTA to 20 ml per liter.

<sup>7</sup>If Sarcosyl powder is added directly to the other components of this reagent, warm the solution to 50- 60°C for 30-60 minutes, or leave at room temperature for ≈2 hours to completely dissolve the Sarcosyl; adjust to the final volume with sterile Ultrapure Water.